

• 论著 •

温下健脾法联合抗菌药物治疗脓毒症的实验研究

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【摘要】目的 探讨温下健脾法协同抗菌药物治疗脓毒症的作用机制。**方法** 选择32只SPF级雄性SD大鼠,采用盲肠结扎穿孔法(CLP)复制大鼠脓毒症模型,制模成功后将大鼠按随机数字表法分为模型组、头孢曲松治疗组、中药治疗组、头孢曲松+中药治疗组,并选择8只SD大鼠作为假手术组。假手术组和模型组给予生理盐水腹腔注射+灌胃;中药治疗组给予生理盐水腹腔注射+大黄附子汤加减方8 mg/kg灌胃;头孢曲松治疗组给予头孢曲松120 mg/kg腹腔注射+生理盐水灌胃;头孢曲松+中药治疗组给予头孢曲松120 mg/kg腹腔注射+大黄附子汤加减方8 mg/kg灌胃,均每日2次,连续给药2 d。给药后观察各组大鼠死亡率;并检测肠道菌群、肠道黏膜通透性[肠道黏膜损伤指数(IMII)、肠道黏膜分泌型免疫球蛋白(sIgA)、血清D-乳酸、二胺氧化酶(DAO)]及血清炎症指标[脂多糖(LPS)、肿瘤坏死因子-α(TNF-α)、白细胞介素-6(IL-6)]水平。**结果** ①大鼠死亡率:头孢曲松+中药治疗组(25.0%)<中药治疗组(37.5%)和头孢曲松治疗组(37.5%)<模型组(50.0%),组间比较差异均有统计学意义(均P<0.05)。②16S rDNA测序分析:模型组大鼠拟杆菌门比例明显低于假手术组[(24.36±7.15)%比(45.20±6.05)%],变形菌门比例明显高于假手术组[(10.03±7.55)%比(0.41±0.21)%];头孢曲松治疗组肠道菌群多样性指数明显低于假手术组及中药治疗组(404.60±17.09比470.80±16.97、469.20±14.59,均P<0.05)。主成分分析(PCA)显示,中药治疗组与假手术组的组成更接近,提示温下健脾法能减少脓毒症大鼠的肠道菌群紊乱。③肠道黏膜病理学改变:光镜下可见假手术组大鼠肠黏膜组织结构完整;模型组小肠黏膜厚度明显变薄,局部可见炎症细胞浸润;中药治疗组、头孢曲松治疗组、头孢曲松+中药治疗组大鼠肠黏膜厚度稍增厚,局部炎症细胞浸润较模型组减少;中药治疗组和头孢曲松+中药治疗组大鼠肠黏膜厚度较头孢曲松治疗组稍增厚,排列规整。④肠道黏膜通透性和炎症状态:模型组IMII、D-乳酸、DAO、LPS、TNF-α和IL-6均较假手术组明显增高[IMII:4.37±0.56比0.26±0.03,D-乳酸(mg/L):12.35±0.83比7.30±1.29,DAO(kU/L):2.16±0.43比0.32±0.06,LPS(kU/L):0.663±0.012比0.095±0.003,TNF-α(μg/L):251.03±82.69比52.15±6.25,IL-6(μg/L):160.50±4.77比54.30±3.36],sIgA较假手术明显降低(μg/L:11.57±0.17比26.76±1.99)。中药治疗组、头孢曲松治疗组、头孢曲松+中药治疗组IMII、D-乳酸、DAO、LPS、TNF-α和IL-6均较模型组降低,sIgA较模型组升高,以头孢曲松+中药治疗组的变化较中药治疗组、头孢曲松治疗组更显著[IMII:1.78±0.23比1.96±0.62、3.39±0.43,D-乳酸(mg/L):8.56±0.37比9.62±0.57、11.42±0.39,DAO(kU/L):1.14±0.12比1.72±0.24、2.01±0.32,sIgA(μg/L):25.34±1.49比23.99±7.85、17.46±1.20,LPS(kU/L):0.302±0.007比0.387±0.004、0.715±0.013,TNF-α(μg/L):57.10±3.98比101.49±21.49、141.91±20.20,IL-6(μg/L):93.71±2.39比87.12±7.31、104.27±1.84]。**结论** 温下健脾法通过调整肠道菌群紊乱,减轻肠道黏膜通透性,降低机体炎症水平,提高抗菌药物治疗脓毒症的作用。

【关键词】 温下健脾法; 脓毒症; 肠道菌群; 肠道黏膜通透性; 炎症水平; 抗菌药物

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【Abstract】Objective To explore the synergism efficacy and mechanism of Warm Purgative and Strengthening Spleen (WPSS) therapy combined with antibiotics in the treatment of sepsis. **Methods** Thirty-two SPF Sprague-Dawley (SD) rats were used to replicate the rat sepsis model by cecum ligation perforation (CLP) method and equally divided into model control (MC) group, ceftriaxone group, Chinese herbal medicine (CHM) group and ceftriaxone + CHM group. Eight SD rats underwent sham surgery were used as a sham operation (Sham) group. Rats in Sham and MC groups were administered with 0.9% normal saline (NS) by intraperitoneal injection and gavage. Rats in CHM group were administered with modified Dahuang Fuzi Decoction (DFD, 8 mg/kg) by gavage + 0.9% NS by intraperitoneal injection, Bid. Rats in ceftriaxone group were administered with 0.9% NS by gavage and ceftriaxone (120 mg/kg) by intraperitoneal injection, Bid. Rats in ceftriaxone + CHM group were administered with modified DFD (8 mg/kg) by gavage and ceftriaxone (120 mg/kg) intraperitoneal injection, Bid. The drugs were given for 2 days. The mortality of rats

in each group was observed after treatment. The intestinal flora changes and intestinal permeability [intestinal mucosa injury index (IMII), intestinal mucosa secretory immunoglobulin (sIgA), serum D-lactic acid, diamine oxidase (DAO) and secretory immunoglobulin (sIgA)] were detected. Meanwhile, the levels of serum inflammation indexes [lipopolysaccharide (LPS), tumor necrosis factor- α (TNF- α) and interleukin-6 (IL-6)] were detected. **Results** ① Mortality: ceftriaxone+CHM group (25.0%) < CHM group (37.5%) and ceftriaxone group (37.5%) < MC group (50.0%), the differences between groups were statistically significant (all $P < 0.05$). ② 16S rDNA sequencing analysis: the ratio of *Bacteroidetes* in MC group was lower than that in the Sham group [(24.36 ± 7.15)% vs. (45.20 ± 6.05)%], and the ratio of *Proteobacteria* in MC group was higher than that in Sham group [(10.03 ± 7.55)% vs. (0.41 ± 0.21)%]. The diversity of intestinal flora in ceftriaxone group was significantly lower than that in Sham and CHM groups (404.60 ± 17.09 vs. 470.80 ± 16.97, 469.20 ± 14.59), the differences between groups were statistically significant (all $P < 0.05$). The principal component analysis (PCA) suggested that the composition of CHM group was closer to that of Sham group, which indicated that WPSS therapy could reduce intestinal flora disorders in rats with sepsis. ③ The pathological changes of intestinal mucosa: light microscope showed the intestinal mucosa of Sham group was intact; the intestinal mucosa became thinner, and local inflammatory cells had infiltration in MC group. The thickness of intestinal mucosa in CHM, ceftriaxone and CHM+ceftriaxone groups was slightly thicker, and the infiltration of local inflammatory cells was less than that in MC group. The thickness of intestinal mucosa in CHM group and ceftriaxone+CHM group was slightly thicker than that in the ceftriaxone group, and the arrangement was more regular than that in MC group and ceftriaxone group. ④ Intestinal mucosa permeability and inflammatory state: IMII, D-lactic acid, DAO, LPS, TNF- α and IL-6 of rats in MC group were higher than those of rats in Sham group [IMII: 4.37 ± 0.56 vs. 0.26 ± 0.29, D-lactic acid (mg/L): 12.35 ± 0.83 vs. 7.30 ± 1.29, DAO (kU/L): 2.16 ± 0.43 vs. 0.32 ± 0.06, LPS (kU/L): 0.663 ± 0.012 vs. 0.095 ± 0.003, TNF- α (μ g/L): 251.03 ± 82.69 vs. 52.15 ± 6.25, IL-6 (μ g/L): 160.50 ± 4.77 vs. 54.30 ± 3.36], while sIgA in MC group was lower than that in Sham group (μ g/L: 11.57 ± 0.17 vs. 26.76 ± 1.99). IMII, D-lactic acid, DAO, LPS, TNF- α and IL-6 of rats in CHM, ceftriaxone and CHM+ceftriaxone groups were significantly lower than those of rats in MC group, while sIgA in CHM, ceftriaxone and CHM+ceftriaxone groups were significantly higher than that of rats in MC group. The change of CHM+ceftriaxone group was more significant than those of CHM group and ceftriaxone group [IMII: 1.78 ± 0.23 vs. 1.96 ± 0.62, 3.39 ± 0.43, D-lactic acid (mg/L): 8.56 ± 0.37 vs. 9.62 ± 0.57, 11.42 ± 0.39, DAO (kU/L): 1.14 ± 0.12 vs. 1.72 ± 0.24, 2.01 ± 0.32, sIgA (μ g/L): 25.34 ± 1.49 vs. 23.99 ± 7.85, 17.46 ± 1.20, LPS (kU/L): 0.302 ± 0.007 vs. 0.387 ± 0.004, 0.715 ± 0.013, TNF- α (μ g/L): 57.10 ± 3.98 vs. 101.49 ± 21.49, 141.91 ± 20.20, IL-6 (μ g/L): 93.71 ± 2.39 vs. 87.12 ± 7.31, 104.27 ± 1.84]. **Conclusion** WPSS therapy may improve the efficacy of antibiotics in the treatment of sepsis by regulating the intestinal flora and reducing the intestinal mucosa permeability and inflammation level.

【Key words】 Warm Purgative and Strengthening Spleen therapy; Sepsis; Intestinal flora; Intestinal permeability; Inflammation level; Antibiotics

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脓毒症是指由感染引起的宿主反应失调导致危及生命的器官功能障碍^[1]。广谱抗菌药物是治疗脓毒症的常用药物,然而大量使用广谱抗菌药物更容易出现细菌耐药、肠道菌群紊乱及肠道屏障损伤^[2]。目前脓毒症的治疗强调采用综合方案,在国内中医药是治疗脓毒症的重要组成部分^[3-4]。中药联合抗菌药物治疗脓毒症具有降低抗菌药物不良反应、提高临床疗效的作用^[5],但具体方法和机制有待进一步研究^[6]。研究表明中医温下健脾法联合抗菌药物可改善脓毒症胃肠功能障碍患者的胃肠功能及预后^[7-8]。基于以上认识,本研究观察温下健脾法联合抗菌药物对脓毒症大鼠胃肠功能的影响,报告如下。

1 材料与方法

1.1 实验动物:选择健康 SPF 级成年雄性 SD 大鼠,

体质量 200 ~ 230 g, 购自北京维通利华实验动物科技有限公司, 许可证号: SCXK(京)2016-0006。

1.2 模型制备方法: 采用盲肠结扎穿孔法(CLP)复制脓毒症大鼠模型^[9], 并加以改进^[10]。

1.3 伦理学: 本实验中动物处置方法符合动物伦理学标准(审批号: 201733)。

1.4 实验分组及给药方法: 将大鼠按随机数字表法分为模型组、头孢曲松治疗组、中药治疗组、头孢曲松 + 中药治疗组, 每组 8 只; 并选择 8 只 SD 大鼠作为假手术组。假手术组大鼠麻醉后仅行开腹手术, 找到盲肠, 但未行结扎或穿孔。制模成功后假手术组和模型组给予生理盐水腹腔注射 + 生理盐水灌胃; 头孢曲松治疗组给予头孢曲松 120 mg/kg 腹腔注射 + 生理盐水灌胃; 中药治疗组给予生理盐水腹

腔注射+大黄附子汤加减方8 mg/kg灌胃,每日2次;头孢曲松+中药治疗组给予头孢曲松120 mg/kg腹腔注射+大黄附子汤加减方8 mg/kg灌胃,每日2次。每组灌胃和腹腔注射的生理盐水的数量相等,共给药2 d。

1.5 检测指标及方法

1.5.1 大鼠死亡率观察:记录制模后及治疗过程中各组大鼠死亡率。

1.5.2 大鼠肠道菌群检测:收集各组大鼠粪便,用液氮快速冷冻,提取基因,采用16S rDNA的方法检测大鼠肠道菌群组成。上海美吉生物医药科技有限公司(中国)利用Miseq PE300平台进行测序。根据16S rDNA的测序结果,使用UPARSE软件version 7.1 <http://drive5.com/uparse/>,根据97%的相似度对序列进行操作分类单位(OTU)聚类;使用UCHIME软件剔除嵌合体;主成分分析(PCA)是一种对数据进行简化分析的技术,这种方法可以有效的找出数据中最主要的元素和结构。利用RDP分类器对每条序列进行物种分类注释。

1.5.3 肠道黏膜通透性测定

1.5.3.1 各组大鼠肠道黏膜病理学改变及肠道黏膜损伤指数(IMII)观察:剪取距盲肠根部小肠2 cm,使用磷酸盐缓冲液(PBS)冲洗干净肠内容物后放入4%多聚甲醛水溶液中固定24 h,按常规方法脱水、浸蜡、石蜡包埋,切片,苏木素-伊红(HE)染色,光镜下观察大鼠肠道黏膜组织病理学改变;根据肠道黏膜病理变化分级^[11]评价IMII。

1.5.3.2 各组大鼠肠道通透性的指标测定:给药结束后腹主动脉取血,离心分装-80℃保存待测。采用酶联免疫吸附试验(ELISA)测定血清D-乳酸和DAO水平。

1.5.3.3 各组大鼠肠道黏膜分泌型免疫球蛋白sIgA水平测定:取大鼠肠道黏膜组织液氮研磨后,取70 mg,加组织裂解液(RIPA)1 mL,超声波破碎30 s,每5 s停止10 s,离心15 min后,取上清液,采用ELISA测定sIgA水平,操作严格按试剂盒说明步骤进行。

1.5.4 各组大鼠炎症因子水平测定:处理大鼠后取全血,分离血清,采用ELISA法测定血清脂多糖(LPS)、肿瘤坏死因子-α(TNF-α)及白细胞介素-6(IL-6)水平。

1.6 统计学方法:使用SPSS 20.0统计软件进行数据分析。符合正态分布的计量资料以均数±标准差($\bar{x} \pm s$)表示,采用单因素方差分析;计数资料以

率表示,采用 χ^2 检验。 $P < 0.05$ 为差异有统计学意义。

2 结 果

2.1 不同处置方法各组大鼠病死率比较:头孢曲松+中药治疗组大鼠死亡率[25.0%(2/8)]<中药治疗组[37.5%(3/8)]和头孢曲松治疗组[37.5%(3/8)]及模型组[50.0%(4/8)],组间比较差异有统计学意义(均 $P < 0.05$)。

2.2 不同处理方法各组大鼠肠道菌群紊乱比较(表1;图1):采用16S rDNA测序分析肠道菌群得到了793个OUT,14个门,22个纲,33个目,57个科,150个属,294个种。各组大鼠肠道菌群厚壁菌门比例比较差异无统计学意义($P > 0.05$),组间大鼠肠道菌群中拟杆菌门和变形菌门比较差异明显,模型组大鼠拟杆菌比例明显低于假手术组,变形杆菌门比例明显高于假手术组,说明脓毒症模型大鼠致病菌增加,条件致病菌下降,存在肠道菌群紊乱。中药治疗组拟杆菌比例高于模型组,而变形杆菌比例低于模型组,中药治疗可以回调脓毒症导致的大鼠肠道拟杆菌门下降和变形杆菌门上升。头孢曲松治疗组变形杆菌门和拟杆菌门比例均明显低于模型组。多样性指数分析显示,模型组多样性指数明显低于假手术组;中药治疗多样性指数明显高于模型组;头孢曲松治疗组多样性指数明显低于假手术组和中药治疗组,提示温下健脾法可能减轻抗菌药物引起的肠道菌群紊乱。PCA属水平结果显示,中药治疗组与假手术组的组成更接近,提示温下健脾法能调节脓毒症大鼠肠道菌群紊乱。以上结果提示温下健脾法可减轻脓毒症及抗菌药物引起的肠道菌群紊乱。

表1 不同处理方法各组大鼠主要肠道菌群组成及多样性比较($\bar{x} \pm s$)

组别	动物数 (只)	厚壁菌门 (%)	拟杆菌门 (%)
假手术组	8	52.91±5.36	45.20±6.05
模型组	4	54.26±26.94	24.36±7.15 ^a
中药治疗组	5	46.61±9.23	45.69±7.98 ^b
头孢曲松治疗组	5	69.13±13.53	27.44±12.22 ^{ac}
头孢曲松+中药治疗组	6	72.00±17.48	17.41±9.45 ^{abc}
组别	动物数 (只)	变形杆菌门 (%)	多样性 指数
假手术组	8	0.41±0.21	470.80±16.97
模型组	4	10.03±7.55 ^a	419.50±31.71 ^a
中药治疗组	5	5.72±4.16 ^{ab}	469.20±14.59 ^b
头孢曲松治疗组	5	2.38±1.61 ^{ab}	404.60±17.09 ^{ac}
头孢曲松+中药治疗组	6	9.36±9.09 ^a	440.70±12.33

注:与假手术组比较,^a $P < 0.05$;与模型组比较,^b $P < 0.05$;与中药治疗组比较,^c $P < 0.05$

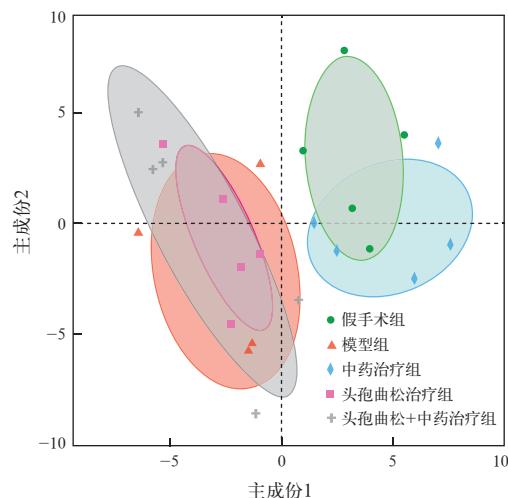


图1 不同处理方法各组大鼠肠道菌群基因水平
主成分分析

2.3 不同处理方法各组大鼠肠道黏膜病理学变化比较(图2):光镜下可见假手术组大鼠肠黏膜组织结构完整,肠绒毛排列整齐;模型组小肠黏膜厚度明显变薄,肠绒毛排列不整齐,部分区域有绒毛上皮剥离,局部可见固有层崩解,绒毛增粗缩短,局部可见炎症细胞浸润;中药治疗组、头孢曲松治疗组、头孢曲松+中药治疗组大鼠肠黏膜厚度稍增加,局部炎症细胞浸润较模型组减少;中药治疗组和头孢曲松+中药治疗组大鼠肠黏膜厚度较头孢曲松治疗组稍增加,排列较模型组和头孢曲松治疗组规整。

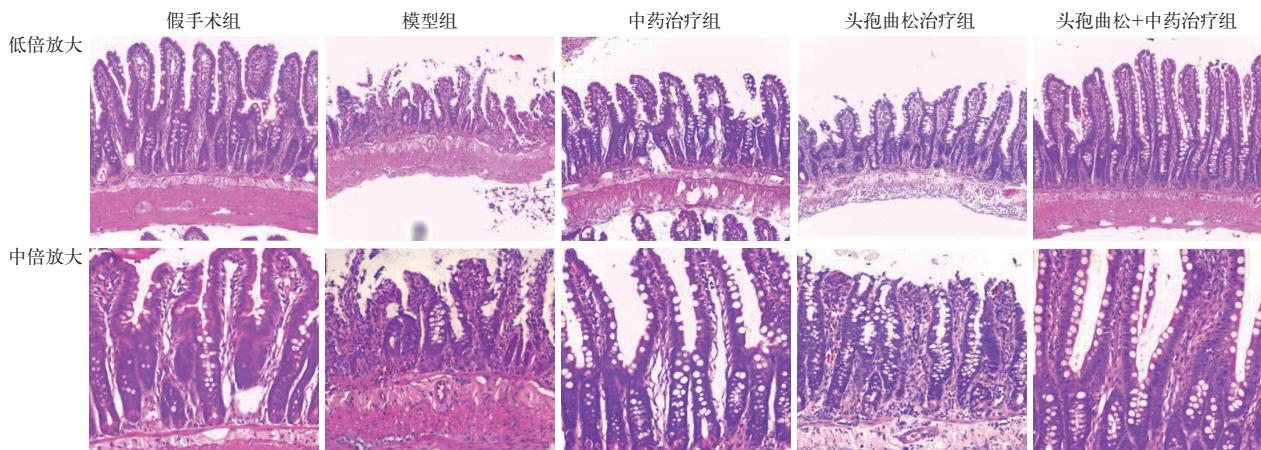


图2 不同处理方法各组大鼠肠道黏膜病理学改变[HE染色 低倍放大(上)和中倍放大(下)]

表2 不同处理方法各组大鼠IMII、血清D-乳酸、DAO和肠黏膜sIgA及炎症指标水平的比较($\bar{x} \pm s$)

组别	动物数 (只)	IMII	D-乳酸 (mg/L)	DAO (kU/L)	sIgA ($\mu\text{g}/\text{L}$)	LPS (kU/L)	TNF- α ($\mu\text{g}/\text{L}$)	IL-6 ($\mu\text{g}/\text{L}$)
假手术组	8	0.26 \pm 0.03	7.30 \pm 1.29	0.32 \pm 0.06	26.76 \pm 1.99	0.095 \pm 0.003	52.15 \pm 6.25	54.30 \pm 3.36
模型组	4	4.37 \pm 0.56 ^a	12.35 \pm 0.83 ^a	2.16 \pm 0.43 ^a	11.57 \pm 0.17 ^a	0.663 \pm 0.012 ^a	251.03 \pm 82.69 ^a	160.50 \pm 4.77 ^a
中药治疗组	5	1.96 \pm 0.62 ^{ab}	9.62 \pm 0.57 ^{ab}	1.72 \pm 0.24 ^{ab}	23.99 \pm 7.85 ^{ab}	0.387 \pm 0.004 ^{ab}	101.49 \pm 21.49 ^{ab}	87.12 \pm 7.31 ^{ab}
头孢曲松治疗组	5	3.39 \pm 0.43 ^{ac}	11.42 \pm 0.39 ^{ac}	2.01 \pm 0.32 ^a	17.46 \pm 1.20 ^{ab}	0.715 \pm 0.013 ^{abc}	141.91 \pm 20.20 ^{abc}	104.27 \pm 1.84 ^{ac}
头孢曲松+中药治疗组	6	1.78 \pm 0.23 ^{abd}	8.56 \pm 0.37 ^{bd}	1.14 \pm 0.12 ^{abd}	25.34 \pm 1.49 ^{bd}	0.302 \pm 0.007 ^{abd}	57.10 \pm 3.98 ^{bd}	93.71 \pm 2.39 ^{abd}

注:与假手术组比较,^aP<0.05;与模型组比较,^bP<0.05;与中药治疗组比较,^cP<0.05;与头孢曲松治疗组比较,^dP<0.05

2.4 不同处理方法各组大鼠IMII、血清D-乳酸、DAO和肠道黏膜sIgA水平的比较(表2):模型组大鼠IMII血清D-乳酸和DAO水平均较假手术组明显升高,肠道黏膜sIgA水平较假手术组明显降低(均P<0.05);中药治疗组及头孢曲松+中药治疗组大鼠IMII、D-乳酸及DAO均较模型组明显降低,sIgA水平明显高于模型组(均P<0.05);头孢曲松+中药治疗组IMII和D-乳酸及DAO均明显低于头孢曲松治疗组,sIgA水平明显高于头孢曲松治疗组(均P<0.05)。

2.5 不同处理方法各组大鼠炎症指标水平比较(表2):模型组LPS、TNF- α 和IL-6水平均明显高于假手术组;中药治疗组、头孢曲松治疗组及头孢曲松+中药治疗组LPS、TNF- α 及IL-6均明显低于模型组,且以头孢曲松+中药治疗组的降低程度更显著(均P<0.05)。

3 讨论

多器官功能障碍是脓毒症患者死亡的重要原因^[12]。早期使用广谱抗菌药物是治疗脓毒症的关键,但同时会造成肠道菌群紊乱。脓毒症肠道菌群紊乱、肠道通透性增加,胃肠功能障碍,条件致病菌入血均可加重脓毒症的炎症状态,进而加剧脓毒症病情的进展^[13-14]。抗菌药物主要抑制或杀灭病原体本身;中药治疗感染性疾病疗效确切,不仅能抑

杀病原体、消除毒素,而且对感染所致炎症、免疫异常、功能紊乱有一定的调节作用,中药与抗菌药物联用具有互补性^[15]。

中医学认为脓毒症患者多因感染、热毒、创伤、手术等导致正气不足,脾胃虚弱,胃失受纳,脾失运化,精微不得化生,气机升降失调。从中药药性理论来看,抗菌药物属于“苦寒”之品,脓毒症患者大量使用抗菌药物及液体复苏往往伤及脾阳,使脾运化水湿功能减弱,痰饮、水湿、瘀血内生,更加阻遏了气机的升降;气机升降失司可导致肠道腑气不畅,变生腹胀、便秘或腹泻,肠鸣音减弱或消失,加剧胃肠功能障碍。通腑攻下法是脓毒症的重要治疗方法^[3, 16],但“寒下”往往加重脾胃损伤。结合中医药防治危重病症时务必顾护脾胃气,恢复脾胃功能的思想,温下健脾法可通腑泻浊、温阳健脾,顾护脾胃,有助于减轻抗菌药物“苦寒”之性,二者联合有助于提高疗效。本研究采用经典温下健脾方剂大黄附子汤加用健脾中药人参、茯苓、生黄芪、甘草等药物,大黄具有清热通腑、活血解毒之功效,附子大辛大热,温阳祛寒,大黄藉附子之大热,其寒性去而走泻之性得存,具有通腑活血解毒功效;细辛辛散温通,可宣通阳气,散肌表之毒邪;人参、茯苓、生黄芪、甘草健脾益气,全方具有温下健脾之功效。研究表明,温下健脾法可调节脓毒症肠道菌群紊乱、改善肠道屏障,联合抗菌药物治疗脓毒症可以改善脓毒症预后及胃肠道功能^[17-18]。

基于以上认识,本研究将脓毒症大鼠分为模型组、中药治疗组、头孢曲松治疗组及头孢曲松+中药治疗组,从肠道菌群-肠道通透性-炎症水平探讨温下健脾法对抗菌药物治疗脓毒症的增效机制。结果表明,中药治疗组、头孢曲松治疗组及头孢曲松+中药治疗组均可降低脓毒症大鼠的死亡率;与单纯头孢曲松治疗组或中药治疗组比较,头孢曲松+中药治疗可降低脓毒症大鼠的病死率。肠屏障是指肠道能防止肠腔内的有害物质通过肠道黏膜进入体内的总和。研究表明,脓毒症及头孢曲松钠使菌群紊乱进一步加剧,肠道内定植的肠杆菌以及益生菌(乳酸杆菌)大量减少,肠球菌增加,球杆菌比值严重倒置,破坏了肠道内的微生态环境^[19-20]。本研究显示,温下健脾法能减轻脓毒症及头孢曲松引起的肠道菌群紊乱。D-乳酸是存在于肠道中细菌的代谢产物。DAO在小肠黏膜或纤毛上皮细胞中表达。当肠道黏膜损伤,通透性增高时,D-乳酸及DAO释放进入循环,血清D-乳酸及DAO升高。

肠道黏膜损伤、血清D-乳酸及DAO水平可反映肠道通透性和损伤^[21],脓毒症肠道黏膜sIgA表达水平降低,增加了肠道的通透性^[22]。本研究结果表明,温下健脾法联合抗菌药物可减轻肠道病理学改变,降低IMII、血清D-乳酸和DAO水平,改善肠道黏膜sIgA的表达。肠道菌群紊乱、肠道通透性增加,条件致病菌进入血中可加重脓毒症的炎症状态,LPS及炎症因子TNF- α 和IL-6在脓毒症患者血清中明显升高^[23]。本研究表明,头孢曲松+中药治疗组血清LPS及TNF- α 和IL-6的水平明显低于单纯头孢曲松治疗组。表明温下健脾法可以调节肠道菌群,减轻肠道黏膜的通透性,降低机体炎症水平,增加抗菌药物治疗脓毒症的作用。同时本研究的不足之处是未能明确温下健脾中药哪些成分、具体如何调节炎症及肠道屏障,这些问题有待进一步研究。

综上所述,抗菌药物是早期治疗脓毒症的关键,温下健脾法联合抗菌药物治疗脓毒症的作用优于单纯抗菌药物,其机制可能是通过调节抗菌药物治疗脓毒症引起的肠道菌群紊乱,减轻肠道黏膜通透性,降低机体炎症反应水平,而发挥与抗菌药物的增效作用,为中西医结合治疗脓毒症提供一定的理论依据。

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• 学术活动预告 •

第七次世界中国中西医结合大会

由中国中西医结合学会、山东中医药大学主办,山东中西医结合学会、山东中医药大学附属医院承办的第七次世界中西医结合大会定于2019年12月5—8日在济南市山东大厦召开,本次大会主题是“中西医结合传承发展,保障人类健康”。

会议将邀请中西医结合相关领域的两院院士、国内外著名专家学者就中西医结合医学的发展、研究成果、创新领域做特邀报告和专题报告,致力于打造一个最具影响力的中西医结合学术交流与知识共享的平台,推动世界中西医结合事业的繁荣与发展。大会设10个分会场,分别从麻醉学、眼科、妇产科、骨伤科、消化内镜、疼痛学、科研院所、眩晕病、风湿类疾病、急救医学等多个专业领域开展学术活动,涉及①中西医结合理论研究;②近五年来中西医结合医学在临床研究、基础研究、药学研究、教学研究、学科建设、政策研究等方面取得的重要成果和宝贵经验;③各临床专业学科新诊疗经验的总结和分析,中西医结合新技术、新方法的推介与评价,以及实现科研成果向临床应用转化的新经验与新模式;④中西医结合优势病种临床诊疗路径的实践经验与临床共性问题的探讨,中西医结合标准化研究;⑤中西医结合医学未来的合作与研究领域;⑥中草药药效物质基础研究与开发等专题。

会议时间:2019年12月5—8日。2019年12月5日8:00—13:00报到;2019年12月6—7日8:00—18:00第七次世界中西医结合大会。

会议地点:山东济南山东大厦(地址:山东省济南市中区马鞍山路2-1号,电话:020-87379988)

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