

• 论著 •

西红花酸对百草枯中毒大鼠的肝保护效应

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【摘要】目的 探讨西红花酸对百草枯(PQ)中毒大鼠急性肝损伤的保护机制。**方法** 按随机数字表法将54只雄性Wistar大鼠分为对照组、染毒组和治疗组,每组再分为染毒后0.5、2、6d亚组($n=6$)。采用腹腔注射20%PQ 20 mg/kg制备PQ中毒急性肝衰竭模型;对照组注射等量生理盐水。治疗组0.5d后腹腔注射西红花酸50 mg/kg,每日1次,直至处死动物;另两组注射等量生理盐水。各组制模后于相应时间点处死大鼠,收集下腔静脉血和肝组织。苏木素-伊红(HE)染色后光镜下观察6d肝组织病理学改变;采用酶联免疫吸附试验(ELISA)检测血清肿瘤坏死因子- α (TNF- α)、白细胞介素-6(IL-6)水平;采用反转录-聚合酶链反应(RT-PCR)检测诱导型一氧化氮合酶(iNOS)、核转录因子- κ B(NF- κ B)的mRNA表达;采用底物显色法检测6d肝组织凋亡相关因子天冬氨酸特异性半胱氨酸蛋白酶(caspase-8、-9、-12)活性。**结果** 染毒组肝组织炎性细胞广泛浸润,弥漫性碎片状坏死,肝细胞再生不明显,且随时间延长而加重;治疗组肝小叶结构尚存,可见点状坏死、少量充血和炎性细胞浸润。染毒组和治疗组0.5、2、6d血清IL-6、TNF- α 水平和肝组织iNOS、NF- κ B的mRNA表达,以及6dcaspase-8、-9、-12活性均较对照组显著升高;而治疗组各指标则较染毒组显著降低[IL-6(ng/L):0.5d为188.37±64.21比376.61±82.42,2d为287.18±58.69比432.77±96.28,6d为234.24±10.17比375.41±37.59;TNF- α (ng/L):0.5d为472.36±76.43比688.33±102.19,2d为189.32±87.54比296.21±89.77,6d为99.28±16.13比168.41±66.78;iNOS mRNA(灰度值):0.5d为2.998±0.801比3.453±0.026,2d为3.126±0.306比5.259±0.153,6d为0.841±0.135比1.225±0.057;NF- κ B mRNA(灰度值):0.5d为1.569±0.818比2.361±0.063,2d为2.345±0.489比4.668±0.368,6d为2.348±0.316比3.972±0.449;caspase-8(pmol/mg):6d为126.77±9.97比199.18±66.48;caspase-9(pmol/mg):6d为213.12±69.06比321.62±89.39;caspase-12(pmol/mg):6d为183.46±70.52比219.68±53.93,均 $P<0.05$]。**结论** 西西红花酸对PQ中毒大鼠具有肝保护效应,其作用可能是通过降低血中炎性因子水平,抑制肝组织caspase-8、-9、-12活性及iNOS、NF- κ B基因表达来实现的。

【关键词】 中毒; 百草枯; 西西红花酸; 肝损伤; 保护效应; 核转录因子- κ B

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Liver protection of crocetin against paraquat poisoning in rats Gao Ke, Guo Hongxing, Liu Liangming, Ding Yanqing, Kuang Meile, Li Jisheng

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【Abstract】Objective To study the liver protection of crocetin against paraquat (PQ) poisoning induced acute liver injury in rats. **Methods** Fifty-four male Wistar rats were randomly divided into control group, exposure group and treatment group, and the rats in each group were subdivided into the 0.5th, 2nd, and 6th day after exposure subgroups ($n=6$). The model of acute liver failure induced by PQ poisoning was reproduced by intraperitoneal injection of 20 mg/kg of 20% PQ, and the rats in control group was injected with the same amount of normal saline. The rats in treatment group were given with intraperitoneal injection of 50 mg/kg crocetin after 0.5 day, once a day until they were sacrificed; the other two groups were injected with the same amount of normal saline. The rats in all groups were sacrificed at the corresponding time points, and blood was collected from inferior vena cava and hepatic tissue was

harvested. Hematoxylin and eosin (HE) staining was used to observe the pathological changes in liver tissue on the 6th day under light microscope. Enzyme linked immunosorbent assay (ELISA) was used to detect the serum tumor necrosis factor- α (TNF- α) and interleukin-6 (IL-6) levels. Reverse transcription-polymerase chain reaction (RT-PCR) was used to detect the mRNA expressions of inducible nitric oxide synthase (iNOS) and nuclear factor- κ B (NF- κ B). The activities of apoptosis related factors, including caspase-8, -9, -12, in hepatic tissue were determined on the 6th day with chromogenic substrate method. **Results** In the liver tissue of exposed group, extensive infiltration of the inflammatory cells and the diffuse fragments necrosis were visible, and the regeneration of the liver cells was not obvious, and severity of the injury in a time dependent way. In the treatment group, the structure of hepatic artery was visible, and the infiltration of necrosis, congestion and inflammatory cells were not obvious. On the 0.5th, 2nd, and 6th day, serum levels of IL-6 and TNF- α , the mRNA expressions of iNOS and NF- κ B in liver tissue, and the caspase-8, -9, -12 activities on the 6th day in the exposure group and treatment group were significantly higher than those in the control group. And the parameters in treatment group were significantly lower than those of the exposure group [IL-6 (ng/L): 188.37 ± 64.21 vs. 376.61 ± 82.42 on the 0.5th day, 287.18 ± 58.69 vs. 432.77 ± 96.28 on the 2nd day, 234.24 ± 10.17 vs. 375.41 ± 37.59 on the 6th day; TNF- α (ng/L): 472.36 ± 76.43 vs. 688.33 ± 102.19 on the 0.5th day, 189.32 ± 87.54 vs. 296.21 ± 89.77 on the 2nd day, 99.28 ± 16.13 vs. 168.41 ± 66.78 on the 6th day; iNOS mRNA (gray value): 2.998 ± 0.801 vs. 3.453 ± 0.026 on the 0.5th day, 3.126 ± 0.306 vs. 5.259 ± 0.153 on the 2nd day, 0.841 ± 0.135 vs. 1.225 ± 0.057 on the 6th day; NF- κ B mRNA (gray value): 1.569 ± 0.818 vs. 2.361 ± 0.063 on the 0.5th day, 2.345 ± 0.489 vs. 4.668 ± 0.368 on the 2nd day, 2.348 ± 0.316 vs. 3.972 ± 0.449 on the 6th day; caspase-8 (pmol/mg): 126.77 ± 9.97 vs. 199.18 ± 66.48 on the 6th day; caspase-9 (pmol/mg): 213.12 ± 69.06 vs. 321.62 ± 89.39 on the 6th day; caspase-12 (pmol/mg): 183.46 ± 70.52 vs. 219.68 ± 53.93 on the 6th day, all $P < 0.05$]. **Conclusion** Crocetin has protective effect on liver in rats with PQ poisoning, which role is related with reducing the blood level of inflammatory factors, inhibiting the hepatic caspase-8, -9, -12 activities and gene expressions of iNOS and NF- κ B.

【Key words】 Poisoning; Paraquat; Crocetin; Liver injury; Protective effect; Nuclear factor- κ B

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百草枯(PQ)毒性极大,且无特效解毒药,口服中毒病死率高达90%以上,早期对肝脏损伤极大。西红花是鸢尾科番红花属球根类草本植物,又称番红花和藏红花。西红花酸是西红花的主要有效成分之一,具有多不饱和共轭烯酸结构,属类胡萝卜素物质。研究表明西红花酸具有多种生物功能,能有效促进细胞生长和抑制细胞凋亡^[1]。本研究通过建立PQ中毒急性肝衰竭动物模型,探讨西红花酸对肝脏的保护效应,为今后选择有效的中医中药治疗方法提供实验依据。

1 材料与方法

1.1 实验材料: 20% PQ购自浙江升华拜克生物股份有限公司;西红花酸购自美国Sigma公司。肿瘤坏死因子- α (TNF- α)、白细胞介素-6(IL-6)酶联免疫吸附试验(ELISA)试剂盒购自美国RapidBio公司;反转录-聚合酶链反应(RT-PCR)检测试剂盒、莫洛尼鼠白血病病毒(M-MLV)反转录酶试剂盒购自美国Promega公司;天冬氨酸特异性半胱氨酸蛋白酶(caspase-8、-9、-12)活性检测试剂盒购自美国BioVision公司。

1.2 动物分组及处理: 7~9周龄健康雄性Wistar大鼠54只,体重(210 ± 20)g,购自南方医科大学实验动物中心,动物合格证号:SCXK(粤)2011-0015。

按随机数字表法将大鼠分为对照组、染毒组和治疗组,每组再分为染毒后0.5、2、6d亚组,每个亚组6只。采用腹腔注射20%PQ 20mg/kg制备PQ中毒急性肝衰竭模型;对照组注射等量生理盐水。治疗组制模0.5d后腹腔注射西红花酸50mg/kg,每日1次,直至处死;另两组注射等量生理盐水。

本实验经医院伦理委员会批准,动物处置方法符合动物伦理学标准。

1.3 检测指标及方法: 各时间点麻醉处死动物,取下腔静脉血,同时取肝脏组织液氮中保存备检。

1.3.1 ELISA测定血清炎性因子IL-6和TNF- α 水平: 静脉血离心取血清,按照ELISA检测试剂盒说明书步骤操作,以分光光度计检测样品吸光度(A)值,并在标准曲线上获得IL-6、TNF- α 水平。

1.3.2 RT-PCR检测诱导型一氧化氮合酶(iNOS)、核转录因子- κ B(NF- κ B)的mRNA表达: 用TRIzol试剂提取肝组织总RNA。模板用2mg总RNA,合成第一链cDNA,反转录。PCR引物设计采用Primer Premier 5.0软件完成,所得PCR产物经琼脂糖凝胶电泳,采用Bio-Rad Quantity-one 4.7成像软件检测并计算待测基因灰度值,以目的基因与内参照 β -肌动蛋白(β -actin)的灰度值比值作为样品的表达量。

1.3.3 肝组织凋亡相关分子 caspase 活性检测: 取6 d液氮冻存肝组织标本, 经裂解液匀浆处理后, 4℃离心取上清液。采用底物显色法, 用荧光分光光度计分析 caspase-8、-9、-12 的活性。

1.3.4 肝组织病理学观察: 取6 d肝组织, 甲醛溶液固定, 经石蜡包埋、切片、苏木素-伊红(HE)染色后光镜下观察。

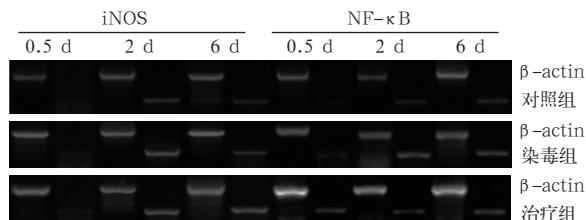
1.4 统计学方法: 采用 SPSS 11.5 软件进行统计学分析, 符合正态分布的计量数据用均数±标准差($\bar{x} \pm s$)表示, 组间比较用方差分析, $P < 0.05$ 为差异有统计学意义。

2 结 果

2.1 血清 IL-6、TNF- α 水平(表 1): 染毒组各时间点血清 IL-6、TNF- α 水平均较对照组显著升高, 而治疗组则较染毒组显著降低(均 $P < 0.05$)。提示西红花酸能减轻 PQ 诱导大鼠炎症反应。

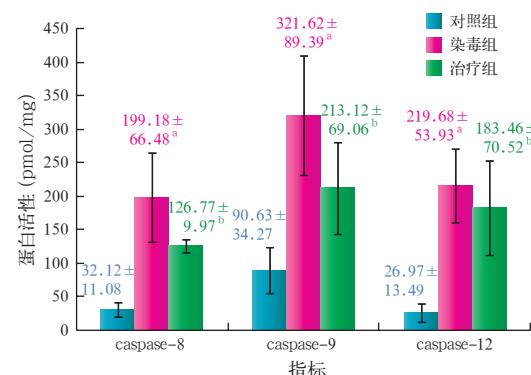
2.2 肝组织 iNOS、NF- κ B mRNA 表达(表 2; 图 1): 染毒组肝组织 iNOS、NF- κ B 的 mRNA 表达均于 2 d 达高峰, 且各时间点均较对照组显著升高; 而治疗组则较染毒组显著降低(均 $P < 0.05$)。提示西红花酸能抑制 PQ 诱导大鼠肝组织 iNOS、NF- κ B 的基因表达。

2.3 肝组织 caspase 活性(图 2): 染毒组 6 d 肝组织凋亡相关因子 caspase-8、-9、-12 活性显著高于对照组, 而治疗组较染毒组显著降低(均 $P < 0.05$)。提示西红花酸能抑制 PQ 诱导大鼠肝细胞凋亡。



RT-PCR 为反转录-聚合酶链反应, iNOS 为诱导型一氧化氮合酶, NF- κ B 为核转录因子- κ B, β -actin 为 β -肌动蛋白

图 1 RT-PCR 检测各组百草枯(PQ)中毒急性肝衰竭大鼠不同时间点肝组织 iNOS 和 NF- κ B 的 mRNA 表达



注: caspase 为天冬氨酸特异性半胱氨酸蛋白酶; 与对照组比较, ^a $P < 0.05$; 与染毒组比较, ^b $P < 0.05$

图 2 西红花酸对百草枯(PQ)中毒急性肝衰竭大鼠 6 d 肝组织 caspase 活性的影响

2.4 肝组织病理学改变(图 3): 对照组肝组织结构正常; 染毒组肝细胞大量空泡状变性, 出现大片状坏死, 小叶结构紊乱, 肝窦充血并出血, 小叶内及汇

表 1 西红花酸对百草枯(PQ)中毒急性肝衰竭大鼠不同时间点血清 IL-6、TNF- α 水平的影响($\bar{x} \pm s$)

组别	动物数 (只)	IL-6(ng/L)			TNF- α (ng/L)		
		0.5 d	2 d	6 d	0.5 d	2 d	6 d
对照组	6	0.14 ± 0.03	1.31 ± 0.17	0.98 ± 0.09	0.12 ± 0.06	2.37 ± 0.19	1.31 ± 0.04
染毒组	6	376.61 ± 82.42 ^a	432.77 ± 96.28 ^a	375.41 ± 37.59 ^a	688.33 ± 102.19 ^a	296.21 ± 89.77 ^a	168.41 ± 66.78 ^a
治疗组	6	188.37 ± 64.21 ^{ab}	287.18 ± 58.69 ^{ab}	234.24 ± 10.17 ^{ab}	472.36 ± 76.43 ^{ab}	189.32 ± 87.54 ^{ab}	99.28 ± 16.13 ^{ab}

注: IL-6 为白细胞介素-6, TNF- α 为肿瘤坏死因子- α ; 与对照组比较, ^a $P < 0.05$; 与染毒组比较, ^b $P < 0.05$

表 2 西红花酸对百草枯(PQ)中毒急性肝衰竭大鼠不同时间点肝组织 iNOS、NF- κ B mRNA 表达的影响($\bar{x} \pm s$)

组别	动物数 (只)	iNOS mRNA(灰度值)			NF- κ B mRNA(灰度值)		
		0.5 d	2 d	6 d	0.5 d	2 d	6 d
对照组	6	0.011 ± 0.007	0.013 ± 0.012	0.013 ± 0.007	0.327 ± 0.043	0.297 ± 0.038	0.305 ± 0.015
染毒组	6	3.453 ± 0.026 ^a	5.259 ± 0.153 ^{ac}	1.225 ± 0.057 ^{ad}	2.361 ± 0.063 ^a	4.668 ± 0.368 ^{ac}	3.972 ± 0.449 ^{ad}
治疗组	6	2.998 ± 0.801 ^{ab}	3.126 ± 0.306 ^{ab}	0.841 ± 0.135 ^{ab}	1.569 ± 0.818 ^{ab}	2.345 ± 0.489 ^{ab}	2.348 ± 0.316 ^{ab}

注: iNOS 为诱导型一氧化氮合酶, NF- κ B 为核转录因子- κ B; 与对照组比较, ^a $P < 0.05$; 与染毒组比较, ^b $P < 0.05$; 与本组 0.5 d 比较, ^c $P < 0.05$; 与本组 2 d 比较, ^d $P < 0.05$

管区炎性细胞浸润,肝细胞再生不明显;治疗组肝小叶结构尚存,肝细胞变性、坏死及肿胀均不明显,可见肝细胞点状坏死,有少量充血和炎性细胞浸润。

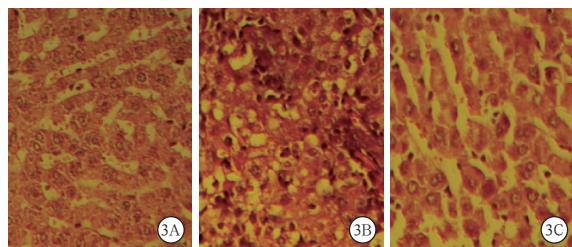


图3 光镜下观察各组大鼠百草枯(PQ)染毒6 d肝组织病理学改变 对照组(A)肝小叶结构正常;PQ染毒组(B)肝组织大片坏死,有大量炎性细胞浸润,库普弗细胞增生肥大;西红花酸治疗组(C)肝小叶结构尚存,坏死及炎性细胞浸润减少 HE染色 中倍放大

3 讨 论

PQ中毒可致多器官功能障碍综合征(MODS),肺是主要靶器官,目前研究最多的是对肝脏的损害作用^[2-4],而其早期对肝脏的损伤亦不容忽视^[5-6]。细胞凋亡是急性肝衰竭最重要的肝细胞死亡形式,研究显示,在疾病高峰期约有70%的肝细胞发生凋亡^[7-9]。本实验探讨了西红花酸对PQ诱导肝细胞凋亡和炎性因子表达的影响,以期为中医中药对PQ中毒致肝衰竭提供有效的治疗方法。

TNF- α 和IL-6等炎性因子与MODS关系最为密切,在炎症发生发展过程中发挥着至关重要的作用^[10-11]。PQ刺激了肝库普弗细胞Toll样受体4(TLR4),引起TNF- α 、IL-6等炎性介质早期大量表达,同时诱导TNF- α 、IL-1 β 、IL-6等细胞因子的基因表达和蛋白质合成^[12-13];TNF- α 可以加重肝微循环障碍,使肝组织发生缺血缺氧性损伤,同时TNF- α 在体内还能刺激IL-6、IL-1 β 的产生^[14-15]。本实验结果提示,西红花酸能抑制PQ诱导大鼠血清TNF- α 、IL-6水平急剧增加,从而显著减轻肝组织的炎症反应。

NF- κ B激活与肝损伤关系密切,其可促进炎性因子大量释放,发挥促炎作用,使肝脏损伤越来越严重^[16];同时,NF- κ B活化可激活IL-6、IL-8等细胞因子合成^[17],放大炎症反应,使肝损伤进一步加重。此外,NF- κ B还参与了iNOS的基因转录,使一氧化氮(NO)含量增加;PQ还可直接通过分泌的TNF- α 诱导iNOS表达,进而引起NO大量合成,NO自由基通过刺激细胞,诱导受感染细胞发生炎症

反应、凋亡和坏死,起到对抗外源性微生物的细胞毒作用^[18];NO自由基还可通过促进周边非损伤细胞的炎症或细胞毒性,加剧超敏反应、脓毒症及自身免疫的组织损伤^[19-20]。本研究提示,西红花酸能极大程度地抑制PQ诱导大鼠肝组织iNOS、NF- κ B的mRNA高表达,从而阻止肝细胞凋亡和炎症损伤。

Caspase活化在细胞凋亡的中心控制和效应阶段起到重要作用^[21]。通常caspase-8、-10、-12介导死亡受体通路的细胞凋亡,分别被募集到Fas和肿瘤坏死因子受体-1(TNFR1)死亡受体复合物;而caspase-9参与线粒体通路的细胞凋亡,被募集到细胞色素C氧化酶/三磷酸脱氧腺苷/凋亡激活因子-1(Cyt C/dATP/Apaf-1)组成的凋亡体(apoptosome),启动caspase活化并开启细胞内的死亡程序,通过异源活化方式水解下游caspase,将凋亡信号放大,同时将死亡信号向下传递。Caspase可由iNOS表达的下游信号分子Bax和NO通过细胞凋亡信号转导而活化^[22-23]。本实验结果提示,西红花酸能够抑制PQ诱导的肝组织caspase-8、-9、-12活性,从而极大程度地减少肝细胞凋亡。

综上所述,西红花酸能显著减少PQ中毒大鼠血清TNF- α 、IL-6的释放,抑制肝组织iNOS、NF- κ B的基因表达及caspase-8、-9、-12活性,从而对肝组织起到保护作用,为临床中医中药治疗PQ中毒提供了新的治疗方法和实验依据。

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